Quality Management of Pre- and Post-Analytical Processes in Laboratory Medicine

Ron B. Schifman, MD Department of Clinical Pathology Tucson Veterans Affairs Medical Center Tucson, Arizona

Abstract: Quality assurance activities in laboratory medicine have traditionally focused on monitoring analytical performance. The scope of quality practices is undergoing gradual change that includes expansion toward continuous monitoring and performance improvement of pre- and post-analytical components of the total testing process. This presentation will address emerging quality management principles and procedures in laboratory medicine, emphasizing specimen quality, appropriateness of testing, results utilization, information quality, user perceptions and benchmarking.

Introduction

Quality management is a vital administrative function that serves to improve performance and add value to products, services and information. Quality management is of considerable value to complex systems such as health care organizations which must integrate widely diverse functions to be efficient and effective. Quality is an attribute that is produced and sustained by making adjustments in a system based on evaluations that come from continuously monitoring performance. 4,5

Quality management in clinical laboratories has focused primarily on following well standardized procedures for maintaining reliable analytic functions. Most quality assessment procedures used in the clinical laboratory today consist of monitoring the accuracy and consistency of reagents, equipment and methods through internal process control, external proficiency testing and on-site inspections. Accrediting organizations and regulatory agencies require adherence to these standardized procedures for laboratory certification and

reimbursement. Analytical process control, while traditionally being the main focus of laboratory quality management, involves only one portion of the total testing process. Concern is growing that a disproportionate amount of time and resources is spent on analytical quality control at the expense of pre- and post-analytical factors that are known to have a considerable impact on the quality of laboratory testing and results utilization.^{7,8} This paper will provide specific examples involving quality management of pre- and post-analytical components of the total testing process.

Specimen quality

The quality of a test result is only as good as the specimen that is submitted for analysis. It is important to continuously examine the quality of specimens that are received and improve processes for optimal specimen collections. Two examples are given describing pre-analytical problems arising from obtaining insufficient number of specimens and from improper timing of collections.

Laboratory diagnosis of tuberculosis

A series of three morning sputum specimens is recommended for mycobacterial culture. Submitting an insufficient number of sputum specimens has been associated with significant delays in diagnosis of pulmonary tuberculosis. 9,10 A College of American Pathologists (CAP) Q-Probes study, conducted in 1994 and involving 534 institutions, disclosed that the median number of specimens collected per patient at each institution was well below 3: 1.8 for inpatients and 1.4 for outpatients. A single positive culture was reported for 17.1 % of patients in whom 2 specimens were collected and for 12.4% of patients in whom 3 specimens were collected. While mycobacterial smear and culture turnaround time has been emphasized as one of the more important indicators of laboratory performance, findings from this study suggest that it is also important to insure that sufficient specimens are collected to achieve optimal test sensitivity.

Therapeutic monitoring of digoxin

Digoxin therapeutic drug monitoring practices were studied in 666 institutions participating in a CAP quality improvement Q-Probes study.¹¹ Of 280,172 digoxin levels studied, 6.7% (n=8,679) were in the toxic range (>2.6 nmol/L). While only 1.6% of specimens were collected inappropriately before steady state had occurred (less than 6 hours after oral dose), 25% of these specimens were in the toxic range. Laboratory policies not requiring the time of the last dose before measurement were associated with higher percentages of specimens drawn before the recommended time had elapsed. This study provides a good example of how improper timing of specimen collections can affect quality

testing. Misinterpreting a falsely elevated digoxin level because of improper specimen collection may affect patient management and has potential for adverse clinical outcome if dosing is inappropriately modified on the basis of erroneous information.

Test utilization

Quality laboratory practices should include processes for improving appropriate test selection and utilization. Examples of quality management challenges described below include processes to control inappropriate test duplication and omissions as well as procedures to improve test selection.

Examination and improvement of test ordering processes using volume indicators

Volume indicator criteria have been used in our laboratory since 1987 to assess and improve processes associated with improper test usage.8 For example, a substantial number of duplicate cholesterol orders were found to be caused by preprinted orders on patients receiving total parenteral nutrition. After reviewing the literature and discussing the indications for this test with clinical colleagues, we deleted these orders from the preprinted forms. A similar solution helped to reduce serum aspartate aminotransferase orders in patients with chest pain who were admitted to the coronary care unit. A substantial volume of duplicate uric acid tests was found to be caused by misinterpreting this test as part of panel because of where it was printed on the physician's order form. Revising this form produced a substantial decline in duplicate uric acid orders (Figure 1).

Weekly uric acid volume 1994 to August 1995

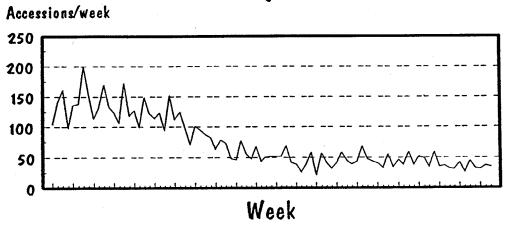


Figure 1. Effect of changing test order form on volume of orders for uric acid

Ova and Parasite Examination on Inpatients

Ova and parasite examinations and bacterial cultures on stool specimens collected from patients who have been hospitalized for 3 or more days are rarely productive. 12-15. In this clinical setting, patients with diarrhea are more likely to have Clostridium difficile infection. Omitting to test for *C. difficile* in hospitalized patients with diarrhea in whom a stool specimen is submitted for ova and parasite examination or bacterial culture may represent poor test selection. When this occurs, it may be necessary to defer testing and consult the physician about indications for evaluating the patient for C. difficile infection (i.e., history of current or recent antibiotic or chemotherapy).

Utilizing of acute viral hepatitis A serology tests

When acute viral hepatitis A is suspected, the infection can be confirmed by measuring IgM specific antibody against hepatitis A antigen (anti-HAV IgM). Since acute viral hepatitis is nearly always associated with elevated of serum aminotransferase (AST or ALT) activity, utilization of anti-HAV (IgM) can be assessed by using the aminotransferase test as an initial indicator of appropriate test selection. In a Q-Probes study involving 625 institutions, the percentage (0.47%) of seropositive anti-HAV (IgM) results observed when aminotransferase results were normal was not significantly different from the percentage (6.27%) of reactive serologic tests reported previously in a healthy population of randomly selected adults.¹⁶ These results show that when accompanied

by normal serum aminotransferase levels, the pretest probability of a positive IgM anti-HAV test is extremely low, and similar to that found in a healthy population. This finding supports a strategy in which serum aminotransferase is used as a prospective utilization review indicator when testing for IgM anti-HAV is ordered. Deferment of serologic testing for acute hepatitis when aminotransferase levels are normal would substantially decrease test volume and improve test selection.

Utilizing Results

One of the most important and challenging quality management goals is to insure that test results are property utilized. A test must be performed correctly and for the proper indication; the results must also be interpreted and applied properly. Failure of physicians to adequately manage patients with low serum vitamin B₁₂ ¹⁷, hypercholesterolemia¹⁸ or anemias¹⁹ are well documented examples of this problem. Methods to insure proper utilization of test results should become an inherent part of clinical laboratory practice.

Utilizing of antimicrobial susceptibility results

When antibiotic resistance is not recognized in a timely fashion, administering appropriate antibiotic therapy may be delayed. Without active review and intervention, the average time lag between susceptibility results reporting and therapeutic modifications is about 24 hours.²⁰ Interestingly, a delayed response to completed results is independent of the speed at which the antimicrobial susceptibility test is performed, even when rapid methods are used.²¹ Patients with serious infections are at risk for delays or failures in treatment, and

given that results from antimicrobial susceptibility tests are predictive of therapeutic responses, unfavorable outcomes. ²²⁻²⁵

We conducted a case-control study that examined the value of correlating therapy with final susceptibility results concurrently, using an integrated computer system.

Among the non-intervention group, no changes were made within 24 hours compared with the intervention group. In the intervention group, an appropriate change in therapy was made in under 24 hours for 54% when a note was written in the patient's chart describing the discrepancy between test results and current antibiotic treatment.

Manufacturers of major automated microbiology systems, having recognized that rapid antimicrobial susceptibility test results alone are insufficient for optimal patient care, are now providing software applications that automatically link pharmacy and microbiology data for review and analysis. This is an important advance in quality management that will enable laboratories to improve their utilization of results.

Telephone results reporting

A Q-Probes study conducted in 1995 evaluated the accuracy of telephone inquires about specimen requirements and test results in 459 institutions. A questionnaire revealed that 39% and 60% of institutions had written guidelines for handling telephone inquires and dealing with security, respectively. Of 5,865 calls made about specimen requirements, 73% were correct, 13.4% were partially correct, 9.6% were incorrect and 3.9% were not completed. Of 2,948 calls made to obtain test results, 3.5% were abandoned. For all completed calls, 2.4%

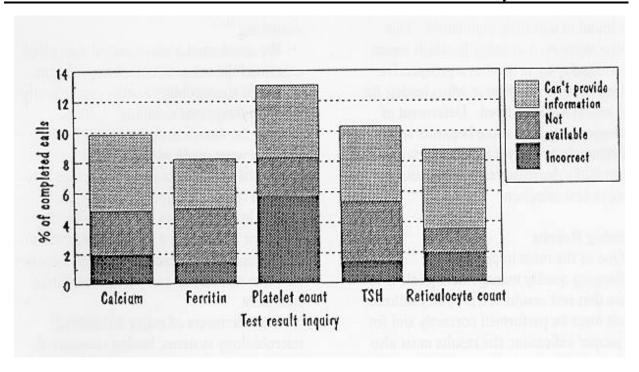


Figure 2. Accuracy of test results reporting by telephone (CAP Q-Probes study)

were incorrect, 2.7% indicated that results were not yet available, and for 4.8% of these, test results could not be given, found, or were unknown (Figure 2). Of 2,806 responses, 23.8% included correct information about tests, and 15.4% indicated that test results were abnormal (all cases selected had test results that exceeded the reference range).

Based on these results we recommended that clinical laboratories: 1) encourage use of computer systems in lieu of telephone support for providing information about test results and specimen requirements, 2) develop standards for telephone support consistent with how information is provided in written and computer formats, 3) always indicate that a test result is abnormal if it is outside the reference range when providing results by telephone and 4) develop written

instructions for employees handling telephone inquires.

Benchmarking

Quality indicators gain substantial value by being interpreted in comparison with a peer group. Q-Probes is a CAP voluntary subscription improvement program for interinstitutional quality assessment and improvement.^{26,27} Participants perform quality assessment studies dealing with many different types of pre- and post-analytical components of the testing process. The data collected by each facility are compared with aggregate data from other institutions as a benchmark to gauge individual performance. A critique is prepared for each study providing an interpretation of the summarized data and suggestions for improvement. While some examples of Q-

Analytical Turnaround Time Antimicrobial Susceptibility Patters Autologous Blood Utilization

Autopsy Contributions in Quality Assurance

Adequacy

Autopsy Report Adequacy

Performance

Autopsy Timeliness and Permit Adequacy

Bedside Glucose Monitoring

Bladder Carcinoma Surgical Pathology Report Adequacy

Blood Culture Contamination Blood Culture Utilization

Blood Bank Control of Usage and Wastage

Breast Carcinoma Surgical Pathology Report Adequacy

Cervical Biopsy - Cytology Correlation Cervico-vaginal Cytology Specimen Adequacy Cervico-vaginal Cytology Specimen Adequacy

Chemistry Specimen Acceptability Coagulation Test Utilization

Colorectal Carcinoma Surgical Pathology Report Adequacy

Complications of Phlebotomy

Critical Values Duplicate Test Orders

Time

Emergency Department Turnaround Time Emergency Department Turnaround Time

Extraneous Tissue

Fine Needle Aspiration Cytohistologic Correlation (FNAC)

Frozen Section Turnaround Time

Handling of Mammographically Detected Breast Biopsy Tissue

Hematology Specimen Acceptability

Inpatient Phlebotomy

Laboratory Safety Practices and Policies Laboratory Computer Availability Laboratory Diagnosis of Tuberculosis

Laboratory Quaity Assurance Programs

Laboratory Proficiency Testing

Lung Carcinoma Surgical Pathology Report

Lung Cancer FNAC Diagnostic

Nosocomial Infection Rates

Order Accuracy

Pap Smear Rescreening

Patient Satisfaction with Phlebotomy Service

Post-analytical QA: Hypercalcemia

QC Exceptions

Quality of Telephone Responsiveness Reference Test Service Quality

Reporting Error

Routine Test Turnaround Time Sputum Specimen Adequacy

Stool Microbiology

Surgical Pathology Specimen Ident & Accessioning Surgical Pathology Frozen Section Consultation Surgical Pathology Complex Spec Turnaround Time Surgical Pathology Routine Biopsy Turnaround

Surgical Pathology Frozen Section Consultations Surgical Pathology Frozen Section Consultations Surgical Pathology Diagnosis Turnaround Time TDM Timing

The INR & Monitoring of Oral Anticoagulants e Timeliness of Urine Specimen Analysis

Transfusion Appropriateness Transfusion Error Reporting

Viral Hepatitis Serology. Test Utilization Wristband Identification Error Reporting

Table 1. Q-Probes Studies 1989 to 1995

Probes studies have already been provided, a complete list of studies between 1989 and 1995 is shown (Table 1).

Conclusion

As can be seen, quality management in clinical laboratories must involve examination of the total testing process. It is necessary to raise expectations and requirements for quality performance beyond analytical process control. Quality assessment and improvement in pre- and

post-analytical phases of testing requires teamwork and inter-departmental cooperation. This brings new challenges as well as opportunities to solve persistent problems and improve the quality of health care.

References

Batalden P.B., E.D. Buchanan.
 Industrial Models of Quality
 Improvement. In: Goldfield N, Nash
 DB, eds. Providing Quality Care.

- Philadelphia, PA: 1989 American College of Physicians; pp. 133-159.
- Berwick DM, Godfrey AB, Roessner J. Curing Health Care. New Strategies for Quality Improvement. A Report on the National Demonstration Project on Quality Improvement in Health Care. 1991, Jossey-Bass, San Francisco, CA.
- 3. Laffel G, Blumenthal D. The case for using industrial quality management science in health care organizations. *JAMA*. 1989;262:2869-73.
- 4. Berwick DM. Continuous improvement as an ideal in health care. *N Engl J Med.* 1989;320:53-6.
- 5. Kritchevsky SB, Simmons BP.
 Continuous quality improvement.
 Concepts and applications for patient care. *JAMA*. 1991;266:1817-23.
- 6. Howanitz PJ, Howanitz JH. The Clinical Laboratory. Wenzel RP (ed). Assessing Quality Health Care. Baltimore, MD: Williams and Wilkins 1992; pp 465-487.
- 7. Bartlett, RC, Mazens-Sullivan M, Tetreault JZ, Lobel S, Nivard J. Evolving approaches to management of quality in clinical microbiology *Clin Microbiol Rev.* 1994;7:55-88.
- 8. Schifman, RB. Quality assurance goals in clinical pathology. *Arch Pathol Lab Med.* 1990;114:1140-4.
- 9. Mathur P, Sacks L, Auten G, Sall R, Levy C, Gordin F. Delayed diagnosis

- of pulmonary tuberculosis in city hospitals. *Arch Intem Med.* 1994;154:306-310.
- 10. Kramer F, Modilevsky T, Valiany AR, Leedom JM, Barnes PF.
 Delayed diagnosis of tuberculosis in patients with human immunodeficiency virus infection.

 Am J Med. 1990; 89:451-6.
- 11. Howanitz PJ, Steindel SJ. Digoxin therapeutic drug monitoring practices: A College of American Pathologists Q-probes study of 666 institutions and 18679 toxic levels. *Arch Pathol Lab Med.* 1993;117:684-690.
- 12. Fan K, Morris AJ, Reller LB. Application of rejection criteria for stool cultures for bacterial enteric pathogens. *J Clin Microbiol*. 1993;31:2233-2235.
- 13. Siegel DL, Edelstein PH, Nachamikin 1. Inappropriate testing for diarrheal diseases in the hospital. *JAMA*. 1990;263:979-982.
- 14. Bowman RA, Bowman JM, Arrow SA, Riley TV. Selective criteria for the microbiological examination of faecal specimens. *J Clin Pathol*. 1992;45:838-39.
- 15. Yannelli BI, Gurevich I, Schoch PE, Cunha BA. Yield of stool cultures, ova and parasite tests, and *Clostridium difficile* determinations in nosocomial diarrheas. *Am J Infect Control*. 1988;16:246-249.

- 16. Jensen DM, Dickerson DD, Linderman MA, Kessler H. Serum alanine aminotransferase levels and prevalence of hepatitis A, B, and delta in outpatients. *Arch Intern Med.* 1987;147:1734-7.
- 17. Carmel R, Denson TA, Mussell B: Anemia:Textbook vs practice. *JAMA*. 1979;242:2295-7.
- 18. Landzert JS, Heim CR. Physician recognition and treatment of hypercholesterolemia. *Arch Intern Med.* 1989;149:933-35.
- 19. Carmel R, Kamaze DS. Physician response to low serum cobalamin levels. *Arch Intern Med*. 1986;146:1161-65.
- 20. VonSeggem RL. Culture and antibiotic monitoring service in a community hospital. *Am J Hosp Pharm.* 1987;44:1358-62.
- 21. Granato PA. The impact of sameday tests versus traditional overnight testing. *Diagn Microbiol Infect Dis*. 1993;16:237-243.
- 22. Campo L, Mylotte JM. The use of microbiology reports by physicians in prescribing antimicrobial agents. *Am J Med Sci.* 1988;296:392-8.

- 23. Pestotnik SL, Evans RS, Burke JP, Gardner RM, Classen DC.
 Therapeutic antibiotic monitoring:
 Surveillance using a computerized expert system. *Am J Med Sci*.
 1990;88:43-8.
- 24. Trenholme GM. Kaplan RL, Karakusis PH, Stine T, Fuhrer J, Landau W, Levin S. Clinical impact of rapid identification and susceptibility testing of bacterial blood culture isolates. *J Clin Microbiol*. 1989;27:1342-45.
- 25. Schentag JJ, Ballow CH, Fritz AL, Paladino JA, Williams JD, Cumbo TJ, Ali RV, Galletta VA, Gutfeld MB, Adelman MH. Changes in antimicrobial agent usage resulting from interactions among clinical pharmacy, the infectious disease division, and the microbiology laboratory. *Diagn Microbiol Infect Dis.* 1993;16:255-264.
- Bachner P, Howanitz PJ. Q-Probes:
 A tool for enhancing your lab's QA.
 Med Lab Observer. 1991;Nov:37-46.
- 27. Howanitz PJ.. Quality assurance measurements in departments of pathology and laboratory medicine. *Arch Pathol Lab Med*. 1990;114:1131-1135.